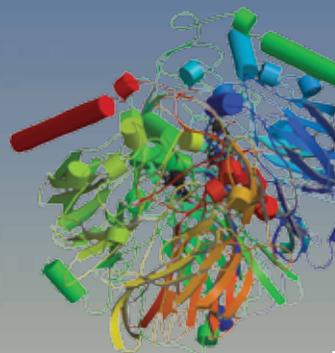


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Keynote Presentations



Blocking Cell Signaling with Recombinant Antibodies

John McCafferty, Ph.D.,
University of Cambridge



BiTE Antibodies in Cancer Therapy

Roman Kischel, M.D.,
Amgen Research
(Munich) GmbH



Protein Expression in Drug Discovery – New Challenges, New Solutions

Lorenz M. Mayr, Ph.D.,
Novartis Pharma



Nanobody Stabilization of G Protein-Coupled Receptor Conformational States: Implications

for Expression
Jan Steyaert, Ph.D., Vrije
Universiteit Brussel

Novel Antibody Constructs and Alternative Scaffolds

Creative Engineering, Delivery Technologies and Targeting

6-7 November

Enhanced Product Properties and Therapeutic Application

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Protein Expression

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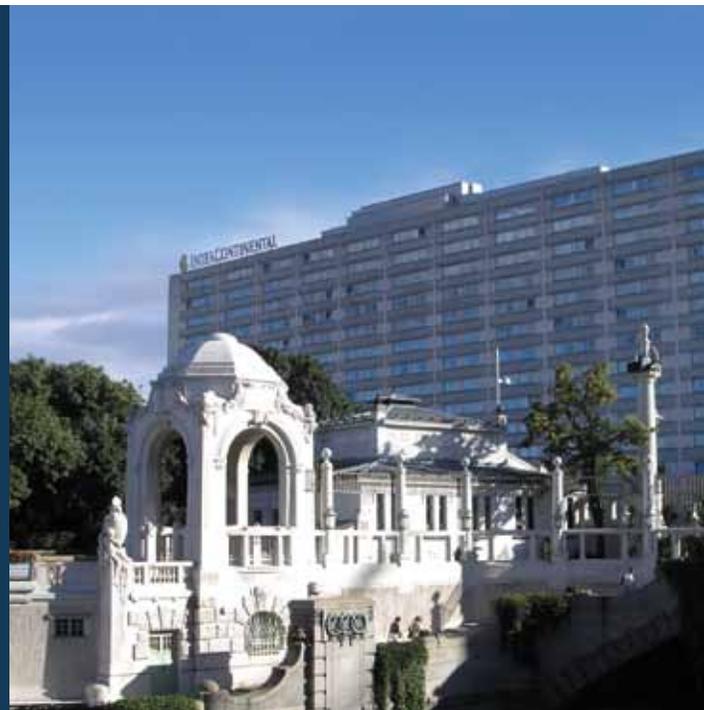
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Pre-Conference Short Courses

Monday, 5 November 2012

SC1: From Understanding of Aggregation to Devising of Prevention Strategies*

9:00 – 10:00 Short Course Registration

10:00 – 13:00 Short Course

Course Instructors:

Martinus Capelle, Ph.D., Senior Scientist and Group Leader, Therapeomic, Inc.

Felix Heise, Ph.D., Senior Scientist and Group Leader, Pharmaceutical and Processing Development, Parenterals Late Stage, F. Hoffmann-La Roche AG

Attend this half-day workshop to gain a critical overview of the available techniques for detection of aggregation and impurities (leachables) and how these methods can be applied. Delegates will learn about strategies for combining analytical methods (e.g. fluorescence spectroscopy and use of fluorescent dyes, field flow fractionation, Nanosight, flow imaging) to ensure detection of aggregates across a range of particle sizes. High throughput analysis (HTA) and high throughput formulation (HTF) platforms will be presented. Using case-studies, potential causes of aggregation and prevention strategies will be discussed.

- Causes and avoidance of impurities and aggregates
- Impact of impurities and leachables
- New technologies for characterization
- Prevention strategies
- Discussion to include experiences regarding aggregation of the participants.

SC2: Measures to Enhance Half-Life and Stability*

13:00 – 14:00 Short Course Registration

14:00 – 17:00 Short Course

Course Instructors:

Arne Skerra, Ph.D., Professor, Chair of Biological Chemistry, Technical University Munich; CEO, XL-protein GmbH

Florian Rueker, Ph.D., Professor, Biotechnology, University of Natural Resources and Life Sciences, Vienna; Co-Founder, f-star

This half-day workshop will provide an overview on the current state of the art in protein engineering, targeted at the improvement of

half-life and stability of proteins, which are key points to consider in the development of biopharmaceuticals. Current technologies to prolong the circulation of biologics, such as PEGylation, fusion with biological polymers (PASylation etc.) and others will be covered and discussed. A range of strategies is available today to engineer the stability of proteins, including targeted mutations and *in vitro* directed evolution. Depending on the fold characteristics (helix-bundles, beta-barrels and the like), single or combined point mutations or additional disulfide bridges can be applied. Experimental methods to identify, engineer and characterize stabilized proteins will be presented in the workshop.

- Management of short plasma half-life, a problem of most biopharmaceuticals
- Current technologies to prolong the circulation of biologics
- PEGylation versus biological polymers (PASylation etc.)
- Aspects of protein scaffold stability (helix-bundles, beta-barrels etc.)

SC3: Engineering of Bi-Specific Antibodies*

13:00 – 14:00 Short Course Registration

14:00 – 17:00 Short Course

Course Instructors:

Julian Bertschinger, Ph.D., CEO, Covagen AG

Francois Rousseau, Head, Antibody Engineering, NovImmune SA

By attending this interactive workshop, you will learn about the various approaches used for the engineering of bi-specific therapeutic antibodies and scaffold-based binding proteins.

The different technologies will be compared, and examples for applications of bi-specific antibodies in drug development will be presented with a focus on bi-specific antibodies that are currently evaluated in clinical trials. Opportunities and challenges in the field of bi-specific antibodies will be discussed.

- Overview on bi-specific antibody technologies
- Past and current challenges for bi-specific antibody development
- Alternative scaffold-based bi-specifics
- Case studies of bi-specific antibodies in clinical development
- Discussion of applications and opportunities for bi-specific antibodies

*Separate Registration Required

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Part One: Creative Engineering, Delivery Technologies and Targeting

6–7 November 2012

TUESDAY, 6 NOVEMBER

7:30 Registration and Morning Coffee

8:30 Chairperson's Opening Remarks

Creative Engineering

8:35 Engineering Bi-Specific Fynomer-Antibody Fusions for Therapeutic Applications

Julian Bertschinger, Ph.D., CEO, Covagen AG

Fynomers are small binding proteins derived from the human Fyn SH3 domain. Here, we describe the engineering and characterization of bi-specific Fynomer-antibody fusions targeting two different epitopes on HER2. These Fynomer-antibody fusions prevent effectively proliferation of HER2 positive tumor cells *in vitro* and *in vivo*. Engineered Fynomer-antibody fusion proteins have optimal physico-chemical and *in vivo* half-life properties, making them attractive as drug candidates to be brought into pre-clinical and clinical development.

9:05 High-Throughput Generation of scFv-Fc Antibodies to Novel, Modified and Difficult Targets

Stefan Dübel, Ph.D., Professor, Biotechnology, Technical University of Braunschweig

Today, the control of the biochemical conditions during a recombinant antibody selection process makes possible the production of human antibodies specific for single post-translational modifications or to antigens in complex membrane preparations. With the technology for the generation of human targeting domains being matured now, it is evident that identification of disease related antigens becomes the limiting factor. Our high-throughput pipeline for the generation of scFv-Fc antibodies can provide lead substances right from the target discovery process.

9:35 A Novel Ligation Technology for Site-Specific Engineering and Labeling of Proteins

Graham Cotton, Ph.D., Senior R&D Group Leader, Almac Group

Increasingly, protein engineering and labeling technologies are being used to develop biotherapeutics with improved performance and to enable new therapeutic approaches to be realized using protein-based drugs. To this end, we have developed a novel protein ligation methodology for the site-specific C-terminal modification of proteins. This high yielding, highly selective technology provides a facile approach for the C-terminal PEGylation of proteins, and has broad applicability for the labeling and engineering of proteins including the development of bi-specific protein therapeutics and molecular imaging agents.

» FEATURED PRESENTATION

10:05 Pre-Clinical Advances with PASylated Bi-Specific Combinations of Peptides, Antibody Fragments, and Anticalins

Arne Skerra, Ph.D., CEO, XL-protein GmbH

PASylation, the genetic fusion with conformationally disordered polypeptide sequences composed of the amino acids Pro, Ala, and Ser, allows the facile functional coupling of two different bioactive proteins or peptides. PASylation provides a solvated random chain with large hydrodynamic volume, thus slowing down kidney filtration by a factor 10 to 100 and paving the way to bi-specific drugs with extended plasma half-life. Compared with PEG, PASylation offers several advantages, in particular biodegradability and one-step biotechnological production.

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

Alternative and Novel Deliveries

11:15 Exploiting the Biophysical Properties of Centyrins: New Therapeutic Targets, Novel Routes of Administration and Valuable Tools for Drug Discovery

Robert Hayes, Ph.D., Vice President and Venture Leader, Centyrex Venture, Janssen Research & Development

Alternative scaffolds represent an emerging class of protein therapeutics. We have designed novel consensus FN3 domains, called Centyrins, which have excellent biophysical properties. We are exploiting the robustness of this platform to develop a series of molecules aimed at broadening the therapeutic applications of biologics to areas such as bi-specific drugs, intracellular inhibitors, and alternative routes of drug delivery. By applying automation and high-throughput selections, these molecules are also being used as tools for large and small molecule drug discovery.

11:45 Topical/Local Administration of Single Chain Fv Antibody Fragments in Clinical Settings

Titus Kretzschmar, Ph.D., CSO, Delenex Therapeutics AG

12:15 Sponsored Presentation (Opportunity available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

12:30 Luncheon Presentations or Lunch on Your Own

(Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

14:15 Chairperson's Remarks

14:20 Engineering and Development of Tri-Specific Molecules with Long Plasma Half-Life for Subcutaneous Administration

Fredrik Frejd, Ph.D., Vice President, Research, Affibody AB

A practical obstacle for self-administered subcutaneous injections is the maximal volume that can be injected subcutaneously, as this limits the amount of drug that can be administered. Here we will present our technology for engineering of a minimal multispecific biological substance to increase the dose after subcutaneous administration, animal results of the use of our broad-species cross-reactivity half-life extension technology, and our work to address the challenge of finding a configuration that binds all three targets optimally.

14:50 Treating Ocular and Pulmonary Diseases with Therapeutic Mirror Proteins

Dana Ault-Riché, Ph.D., CEO, Reflexion Pharmaceuticals, Inc.

Mirror proteins are chemically produced entirely out of D-amino acids, making them highly stable, metabolically inert and non-immunogenic, while retaining antibody-like affinity and specificity. Reflexion uses this unique combination of properties to enable new ocular and pulmonary dosing strategies. Reflexion is developing a VEGF-A antagonist to treat wet age-related macular degeneration and diabetic macular edema by drops and an inhaled VEGF-D antagonist to treat a rare disease called lymphangiomyomatosis (LAM), which primarily afflicts young women and currently has no effective treatment.

» FEATURED PRESENTATION

15:20 Transfer^rin Antibodies into the Brain

Mark S. Dennis, Ph.D., Principal Scientist, Antibody Engineering, Genentech, Inc.

Antibodies have a vast therapeutic potential for treatment of CNS diseases, but their passage into the brain is restricted by the blood-brain barrier (BBB). Our most recent efforts to harness receptor-mediated transcytosis pathways of brain endothelial cells in order to deliver a therapeutically relevant dose of antibody across the BBB will be discussed.

15:50 Refreshment Break in the Exhibit Hall with Poster Viewing

16:30 Sponsored Presentation (Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

17:00 Problem Solving Roundtable Discussions

Table 1: The Quest for New and Better Targets for Bispecifics

Moderator: Rebecca Ashfield, Ph.D., Portfolio and Collaborations Manager, Immunocore Ltd.

Table 2: Engineering of Bispecific Antibodies

Moderators: Julian Bertschinger, Ph.D., CEO, Covagen AG and Gregory Adams, Ph.D., Co-Leader, Developmental Therapeutics Program, Fox Chase Cancer Center

Table 3: Small Immunoglobulin Fragments vs. Alternative Scaffolds

Moderator: Christian Heinis, Ph.D., Laboratory of Therapeutic Proteins and Peptides, Institute of Chemical Sciences and Engineering, Ecole Polytechnique Fédérale de Lausanne (EPFL)

Table 4: Issues to do with Formulation and Delivery

Moderator: Dana Ault-Riché, Ph.D., CEO, Reflexion Pharmaceuticals, Inc.

Table 5: Challenges of Targeting Membrane Spanning Proteins

Moderator: Jonathan Belk, Ph.D., Senior Scientist II, Technology and Platform Development, Adimab, LLC.

Table 6: How to Get What You Want From Antibody Phage Display

Moderators: Stefan Dübel, Ph.D., Professor, Biotechnology, Technical University of Braunschweig and John McCafferty, Ph.D., Research Director, Biochemistry, University of Cambridge

18:00-19:00 Welcome Reception in the Exhibit Hall with Poster Viewing

WEDNESDAY, 7 NOVEMBER

7:30 Breakfast Presentation (Sponsorship Opportunity Available) or Morning Coffee

Focus on Targets and Targeting

8:30 Chairperson's Remarks

8:35 Engineering a Novel Class of Targeted Immunocytokines with Superior Properties Compared to Classical Immunocytokines

Ekkehard Moessner, Ph.D., Group Leader, Protein Engineering, pRED Roche, Glycart AG

A novel class of immunocytokines will be discussed that are based on Fc containing and also on non-Fc containing building blocks. The IL2 component is optimized for improved performance in tumor targeting. Enhancement of *in vivo* efficacy, when combined with ADCC competent antibodies, will be discussed.

9:05 Overcoming the Challenges Associated with Developing Antibodies against the Müllerian Inhibiting Substance Type II Receptor - A Highly Conserved Ovarian Cancer Target

Gregory Adams, Ph.D., Co-Leader, Developmental Therapeutics Program, Fox Chase Cancer Center

We have used a variety of strategies to isolate antibodies against the Müllerian Inhibiting Substance Type II Receptor (MISIR). However, as its ligand binding site is highly conserved, none mediate agonistic signaling and apoptosis. To overcome this, we are using rational design strategies based upon modeling the ligand/receptor interaction to design antibodies. This has resulted in lead antibodies that are focused on the ligand binding site. These rationally-designed antibodies and immunoconjugates based upon earlier phage display-derived antibodies will be discussed.

9:35 Antibody-Like Molecules that Engage Multiple Targets

Peter Kiener, Ph.D., President & CEO, Zyngenia, Inc.

We have generated multi-specific antibodies, termed Zybodyes, that are built around a core scaffold antibody and are able to engage five different targets simultaneously in a coordinated cognate manner. The Zybody format allows us generate new therapeutics with novel pharmacology to improve efficacy whilst still retaining all the desirable CMC, stability and production properties of mAbs.

10:05 Sponsored Presentations (Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

» KEYNOTE PRESENTATION

11:05 Blocking Cell Signaling with Recombinant Antibodies

John McCafferty, Ph.D., Research Director, Biochemistry, University of Cambridge

Cell-based screening assays allow identification of functional antibodies which have utility in the treatment of cancer and autoimmune disease. We have used phage display in conjunction with cell-based assays to generate and epitope map human antibodies which block a number of important cell surface targets. Recombinant antibodies were identified which affect signaling in different ways including ligand neutralization, receptor blocking or inhibition of cell surface proteases.

Targeting Multiple Membrane Spanning Proteins

11:35 Selections with Cells and Membranes Using a Yeast-Based IgG Discovery Platform Enable Identification and Optimization of Human Antibodies against Integral Membrane Proteins

Jonathan Belk, Ph.D., Senior Scientist II, Technology and Platform Development, Adimab, LLC.

Complex protein structure and intimate association with lipids represent some of the many technical challenges when working with recombinant forms of integral membrane proteins. Adimab has employed mammalian cells and membrane preparations to successfully develop efficient and reproducible methods for the yeast-based discovery and optimization of antibodies against membrane proteins.

12:05 A Mammalian Display Platform for High-Throughput Discovery of Well-Behaved Fully Human Antibodies – Novel Applications

Marc van Dijk, Ph.D., CTO, 4-Antibody AG

Our retrocyte display platform utilizes retroviral gene transfer of human antibody genes into mammalian pre-B cells to generate stable high diversity antibody display libraries of full length monoclonal human antibodies. These are screened by FACS to yield high quality, fully natural and well expressed antibodies against any target. Data will be presented on several targets and includes applications for targeted identification of antibodies directed to specific epitopes, cross-reactivity to specific species and more.

12:35 Close of Part One: Creative Engineering, Delivery Technologies & Targeting

Part Two: Enhanced Product Properties and Therapeutic Application

7–8 November 2012

WEDNESDAY, 7 NOVEMBER

12:30 – 14:00 Conference Registration for Part Two

14:00 Chairperson's Opening Remarks

Measures to Improve the Properties of the Product

14:05 PK Modulation of Adnectins: Experiences from Bench to Clinic

Ray Camphausen, Associate Vice President, Protein Design, Adnexus, a Bristol-Myers Squibb R & D Company

Adnectins, derived from the highly stable fibronectin 10th Fn3 domain, are among the most advanced alternative scaffolds, with multiple therapeutics in the clinic. The highly engineerable Adnectin scaffold is amenable to a wide variety of approaches to fine tune functional, biophysical, developability, and pharmacokinetic properties in a therapeutic-dependent manner. Multiple approaches to modulate the pharmacokinetics of Adnectins have been explored focused on either increasing hydrodynamic volume and/or leveraging the FcRn recycling pathway. Data reflecting our observations will be presented.

14:35 Ang-2-VEGF Crossmab, a Novel Bi-Specific Human IgG1 Antibody Blocking VEGF-A and Ang-2 Function with Favorable Stability, Half-Life, and Efficacy

Markus Thomas, Ph.D., Senior Research Scientist, Pharma Research and Early Development (pRED), Discovery Oncology, Roche Diagnostics GmbH

VEGF-A blockade has been validated clinically as a treatment for human cancers. Angiopoietin-2 (Ang-2) expression has been shown to function as a key regulator of tumor angiogenesis. We have generated a bi-specific human IgG1 antibody (CrossMab) blocking VEGF-A and Ang-2 function simultaneously. Our data show that the CrossMab has very good stability, an IgG like half-life in cynomolgus monkey and a favorable safety profile. Additionally it shows favorable efficacy in pre-clinical tumor cancer models, thereby representing a promising therapeutic agent for the therapy of cancer patients.

15:05 Novel *in Silico* Prediction Algorithms for the Design of Stable Biologics

Anne Goupil-Lamy, Principal Scientist, Accelrys



Sponsored by

Understanding the effects of mutation

on protein stability and protein binding affinity is an important component of successful protein design, especially in the area of protein therapeutics. *In silico* approaches to predict the effects of amino acid mutations can be used to guide experimental design and help reduce the cost of bringing therapeutics to market. We have implemented and validated several novel methods for fast computational mutagenesis of proteins to calculate the energy effect of mutation on protein stability, and on protein-protein binding affinity with an optional pH dependency calculation. We will present those methods and applications to antibody design.

15:35 Refreshment Break in the Exhibit Hall with Poster Viewing

16:15 Strategies to Extend the Half-Life of Small Recombinant Protein Therapeutics

Roland Kontermann, Ph.D., Professor, Biomedical Engineering, Institute of Cell Biology and Immunology, University of Stuttgart

Half-life extension strategies are becoming increasingly important to improve pharmacokinetic and pharmacodynamic properties. An overview of the various strategies to extend the half-life of recombinant antibodies as well as results from a comparative study including novel strategies utilizing binding to serum albumin and serum immunoglobulins are presented and discussed.

16:45 Optimization of Bi-Specific DART Proteins for Oncology

Syd Johnson, Ph.D., Vice President, Antibody Engineering, MacroGenics, Inc.

The major challenges to development and effective clinical use of fragment-based bi-specific formats have been manufacturability, stability and pharmacokinetics. We have developed and optimized several formats of our stable dual-affinity retargeting (DART) format that address these issues with an emphasis on oncologic applications. Multiple examples will be presented of half-life extended DART proteins that redirect T-cells to tumor-associated antigens, including CMC strategies and *in vitro* and *in vivo* potency. Challenges of non-clinical toxicology for these highly potent molecules in relevant species will also be discussed.

17:15 Problem Solving Roundtable Discussions

Table 1: Engineering of Bispecific Antibodies

Moderator: Florian Rueker, Ph.D., Professor, Biotechnology, University of Natural Resources and Life Sciences, Vienna; Co-Founder, f-star

Table 2: Challenges of Targeting the Immune Response

Moderator: Rebecca Ashfield, Ph.D., Portfolio and Collaborations Manager, Immunocore Ltd.

Table 3: Targeting Tumor Antigens

Moderator: Gregory Adams, Ph.D., Co-Leader, Developmental Therapeutics Program, Fox Chase Cancer Center

Table 4: Measures to Increase Half-Life and Stability of the Product

Moderator: Roland Kontermann, Ph.D., Professor, Biomedical Engineering, Institute of Cell Biology and Immunology, University of Stuttgart

Table 5: Building Manufacturability into Novel Antibody Formats

Moderator: Hans de Haard, Ph.D., CSO, arGEN-X BV

Table 6: Bi/Multispecific Compound Specific Issues When Moving from Pre-Clinical to Clinical and Beyond

Guy Hermans, Ph.D., Principal Scientist, Ablynx NV

18:15 – 19:15 Reception in the Exhibit Hall with Poster Viewing

THURSDAY, 8 NOVEMBER

7:30 Breakfast Presentation (*Opportunity available, please contact Carol Dinerstein, Dinerstein@healthtech.com*) or **Morning Coffee**

8:30 Chairperson's Remarks

Novel Products in Pre-Clinical Development

8:35 Therapeutic Potential of Bicyclic Peptides

Christian Heinis, Ph.D., Laboratory of Therapeutic Proteins and Peptides, Institute of Chemical Sciences and Engineering, Ecole Polytechnique Fédérale de Lausanne (EPFL)

My laboratory is generating bicyclic peptide ligands with high affinity and specificity for disease targets using a phage display-based approach that I had developed with Sir Greg Winter at the Laboratory of Molecular Biology (LMB) in Cambridge, UK. The bicyclic peptides combine the good binding properties of antibodies with favorable characteristics of small molecules. I will present examples of bicyclic peptides antagonists as well as first *in vivo* data.

9:05 Pre-Clinical Developments for Bi-Specific mAb2s with Optimal Target Selection

Florian Rueker, Ph.D., Professor, Department of Biotechnology, University of Natural Resources and Life Sciences, Vienna; Co-Founder, f-star

This talk will describe the established Fcab technology and outline the process for selection of bi-specific mAb2s of high affinity and the rationale behind the choice of bi-specific targets. Moreover it will discuss novel combinations that provide an effect that is more than additive and how this is achieved. *In vitro* and *in vivo* results will be presented.

9:35 Bispecific Immunotherapeutics Based on the RECRUIT Tandab platform

Eugene Zhukovsky, CSO, Affimed Therapeutics AG

10:05 Sponsored Presentation (Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

11:05 Creating Differentiated Antibody Therapeutics

Hans de Haard, Ph.D., CSO, arGEN-X BV

I will present our pre-clinical stage c-Met antibody program, where unprecedented functional diversity has identified a potentially best-in-class antibody product to treat solid tumors. This was done using our platform which generates fully human antibodies via active immunization of outbred llamas. Universal target epitope coverage allows therapeutic grade antibodies with ultra-high potencies and uniquely differentiated qualities to be consistently identified.

»» FEATURED PRESENTATION

11:35 Differentiating Biologics

H. Kaspar Binz, Ph.D., Vice President & Co-Founder, Lead Optimization, Molecular Partners AG

The powerful DARPin platform enables novel therapeutic concepts in which we can tailor efficacy, PK, and mechanism of action at will, allowing for best- and first-in-class products. Differentiation was the key driver for the generation of such powerful drugs for high medical need indications. We will present the latest clinical data on a best-in-class DARPin therapeutic, and highlight different pre-clinical programs including powerful first-in-class multi-specific drugs.

12:05 Generation of Fully Human Monoclonal Antibodies against G-Protein Coupled Receptors

Urs B. Hagemann, Ph.D., Senior Scientist, Discovery, Affitech Research AS

Our technology has successfully generated antibodies against a number of different G-protein coupled receptors (GPCRs). GPCRs compose the largest family of known proteins, covering more than 2% of the human genome and are implicated in the pathology of both inflammatory conditions and cancers. Despite their validated role as drug targets, development of therapeutics has been very challenging due to their structural conformation and the lack of robust screening systems. *In vitro* and *in vivo* data showing applicability of Affitech's anti-GPCR antibodies for cancer treatment will be presented.

12:35 Luncheon Presentations or Lunch on Your Own

(Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

14:00 Chairperson's Remarks

Novel Products in the Clinic

»» KEYNOTE PRESENTATION

14:05 BiTE Antibodies in Cancer Therapy

Roman Kischel, M.D., Principal Scientist, BiTE Technology, Amgen Research (Munich) GmbH

14:35 IMCgp100: A Bi-Specific TCR Anti-CD3 Fusion for the Treatment of Malignant Melanoma

Rebecca Ashfield, Ph.D., Portfolio and Collaborations Manager, Immunocore Ltd.

ImmTACs are soluble, high affinity T cell Receptors fused to an anti-CD3 scFv domain for re-directed T cell killing of tumors. A key differentiating factor of this technology is the ability to target HLA presented epitopes, allowing selection of tumor specific targets which are not expressed on normal cells. ImmTACs have pico-molar potency and recognise very low levels of target antigen, overcoming HLA down-regulation. The presentation will include preliminary data from Phase I/IIa testing of IMCgp100 in malignant melanoma.

15:05 Antibody Mixtures: From Bench to Clinic

Ivan Horak, M.D., CSO/CMO, Symphogen

Over the last decade we have learned more about primary and acquired resistance to targeted therapies. Targeting two non-overlapping epitopes provide a unique opportunity to eliminate key components of oncogenic addiction. A novel mechanism of action has been documented in the preclinical models and ongoing clinical studies will assess the safety and clinical benefit of antibody mixtures.

15:35 Refreshment Break in the Exhibit Hall with Poster Viewing

16:00 Anticalins: Next Generation Clinical Stage Technology with Applications across Multiple Disease Areas and Targets

Marlon J. Hinner, Ph.D., Head, Molecular Biology and Protein Engineering, Pieris AG

We will provide an overview of the Anticalin drug platform and present pre-clinical and clinical data including favorable results of a first in human Phase I study of PRS-050 (anti-VEGF-A). We discuss how we generate potent, stable Anticalins with drug-like properties and significantly lower COGs compared to mAbs. We will also illustrate the benefits of straightforward Anticalin engineering to identify multi-specific Anticalins across therapeutic areas.

16:30 Nanobodies: Proteins Based on the Smallest Functional Fragments of Heavy Chain Antibodies, in the Clinic

Guy Hermans, Ph.D. Principal Scientist, Ablynx NV

Nanobodies are therapeutic proteins based on the smallest functional fragments of heavy chain antibodies, naturally occurring in Camelidae. The modularity of this technology allows for exquisite control over valency, *in vivo* half-life and effector function, and provides a platform for both single- and multi-specific compounds. Examples spanning preclinical up to end of phase II stages across highly diverse indications will be used to illustrate the ease with which these different goals can be met, without compromising on CMC aspects or stability.

17:00 Close of Conference

Part One: Enhancing Expression and Achieving Higher Throughput 6–7 November 2012

TUESDAY, 6 NOVEMBER

7:30 Registration and Morning Coffee

8:30 Chairperson's Remarks

Solving Problems in Existing Systems

» KEYNOTE PRESENTATION

8:35 Protein Expression in Drug Discovery – New Challenges, New Solutions

Lorenz M. Mayr, Ph.D., Executive Director, Unit Head Biology, Protease Platform, Novartis Pharma

Whereas protein expression has long been viewed as a mature science with no need for further improvement, current trends in drug discovery show an increased demand for fast & efficient production systems for recombinant proteins and protein complexes to cope with the demands for protein in sufficient amounts needed for modern hit discovery (HTS, FBS, structure) and lead optimization in discovery research.

9:05 FreEcoli™: A New Series of *Escherichia coli* Strains for Endotoxin-Free Production of Recombinant Proteins and Plasmid DNA

Uwe Mamat, Ph.D., Research Scientist, Structural Biochemistry, Research Center Borstel

We present here the FreE coli™ set of non-conditional *E. coli* derivatives that lack all outer membrane agonists for hTLR4/MD-2 activation. The FreE coli™ strains entirely lack LPS, yet remain viable despite exclusively elaborating the tetra-acylated, endotoxically inactive lipid A precursor lipid IVA. Consistent with the results that activation of hTLR4/MD-2 signaling by FreE coli™ cells was of several orders of magnitude lower compared with cells of the *E. coli* wild-type strain, heterologous proteins and monomeric, supercoiled plasmid DNAs prepared from different FreE coli™ strains were free of endotoxic activity.

9:35 Host Cell and Expression Engineering for Development of an *E. coli* Ketoreductase Catalyst: Enhancement of Formate Dehydrogenase Activity for Regeneration of NADH

Regina Kratzer, Ph.D., Researcher, Institute of Biotechnology and Biochemical Engineering, Graz University of Technology

An *E. coli* co-expressing *Candida tenuis* xylose reductase and *Candida boidinii* formate dehydrogenase (FDH) was developed. Single expression of the FDH gave an enzyme activity of 400 units/gCDW. Co-expression, however, resulted in an 80% decline in FDH activity. Combined effects from increase in FDH gene copy number, supply of rare tRNAs and dampened expression of the ketoreductase brought up the FDH activity 3-fold.

10:05 Does Metabolism Limit Recombinant Protein Production?

Lars Blank, Ph.D., Chair of Applied Microbiology, RWTH Aachen University

The synthesis of proteins is one of the most energy consuming processes in the cell, with the result that cellular energy supply may become critical. We therefore quantified the impact of recombinant protein production on microbial metabolism. The insights into the operation of metabolism during recombinant protein production might guide the optimization of microbial hosts and fermentation conditions

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

11:15 microRNAs in CHO Cell Culture Technology

Matthias Hackl, Ph.D., Research Assistant, Department of Biotechnology, University of Natural Resources and Applied Life Sciences, Vienna

The main impact of the now published genome, transcriptome and miRNome of CHO cells is that it enables researchers to begin to understand the molecular mechanisms of how these cells perform their tasks of efficient growth, high productivity and safe product quality. The focus of this presentation is how this information has improved our ability to use microRNAs for engineering of CHO cells.

11:45 Recombinant IgAs Expressed in CHO Cells: Bottlenecks of Recombinant Cell Lines and Improved Production Strategies

Renate Kunert, Ph.D., Professor, Department of Biotechnology, University of Natural Resources and Life Sciences, Vienna, Austria

Immunoglobulins of subtype A (IgA) mediate a key role in mucosal immunity and are promising new immunotherapeutic candidates. We established recombinant Chinese hamster ovary (CHO) cells which stably expressed two different IgA1 antibodies under serum-free conditions. In this study, we extensively characterized the low and the high producing cell lines in long-term culture and identified bottlenecks in polypeptide expression and assembly.

12:15 Sponsored Presentation (Opportunity available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

12:30 Luncheon Presentations or Lunch on Your Own

(Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

14:15 Chairperson's Remarks

14:20 Improving Transient Gene Expression in Mammalian Cells: The Case of a Human Lysosomal Enzyme

José Luis Corchero, Ph.D., CIBER de Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN), Barcelona

Transient gene expression (TGE) is a useful, fast method to produce recombinant proteins. This work describes TGE-based production of a human enzyme. Several parameters (cells, vectors, and others influencing cell metabolism) were studied and optimized. The proposed protocol allows producing up to several mg/L of active enzyme (or engineered versions) to be tested *in vitro* or in pre-clinical trials.

14:50 CHO Genome Characterization and Engineering to Improve Protein Synthesis and Secretion

Nicolas Mermod, Ph.D., Professor, Laboratory of Molecular Biotechnology, University of Lausanne

We have determined the genome sequence of a CHO cell line used for pharmaceutical production and of derived producer clones. This allowed the characterization of the alterations of the genomic and transgene sequence of candidate cell lines for screening purposes as well as cell engineering for increased protein secretion and modifications. This presentation will illustrate how a systematic and multi-level approach can be used to improve the expression of pharmaceutical proteins, including difficult-to-express ones.

15:20 Rational Engineering of *Escherichia coli* Strains for Plasmid Biopharmaceutical Manufacturing

Geisa Gonçalves, Ph.D., Researcher, Department of Bioengineering, Instituto Superior Técnico (IST), Lisbon, Portugal

15:50 Refreshment Break in the Exhibit Hall with Poster Viewing

16:30 Sponsored Presentation (*Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com*)

17:00 Problem Solving Roundtable Discussions

Table 7: Expressing Protein Complexes for Drug Discovery

Moderator: Lorenz M. Mayr, Ph.D., Executive Director, Unit Head Biology, Protease Platform, Novartis Pharma

Table 8: Solving Expression Problems in CHO Cells

Moderator: Renate Kunert, Ph.D., Professor, Department of Biotechnology, University of Natural Resources and Life Sciences, Vienna, Austria

Table 9: Troubleshooting Transient Gene Expression in Mammalian Cell Lines

Moderator: José Luis Corchero, Ph.D., CIBER de Bioingeniería, Biomateriales y Nanomedicina (CIBER-BBN), Barcelona

Table 10: Genome Engineering to Improve Expression

Moderator: Nicolas Mermoud, Ph.D., Professor, Laboratory of Molecular Biotechnology, University of Lausanne

Table 11: Cell Free or Not Cell Free: Making the Decision/Transition

Moderator to be Announced

18:00-19:00 Welcome Reception in the Exhibit Hall with Poster Viewing

WEDNESDAY, 7 NOVEMBER

7:30 Breakfast Presentation (*Opportunity available, please contact Carol Dinerstein, Dinerstein@healthtech.com*) **or Morning Coffee**

Novel Systems

8:30 Chairperson's Remarks

8:35 Supplementation of Serum Free Media with HT: Implications for Designing Novel CHO-Based Expression Platforms

Speaker to be Announced, Boehringer Ingelheim Pharma GmbH & Co. KG

9:05 Multitechnique Study on a Recombinantly Produced *Bacillus halodurans*/Laccase and an S-Layer/Laccase Fusion Protein

Uwie B. Sleytr, Ph.D., Head, Department of NanoBiotechnology, University of Natural Resources and Life Science

The laccase of *Bacillus halodurans* C-125 was immobilized on the S-layer lattice formed by SbpA of *Lysinibacillus sphaericus* CCM 2177 either by (i) covalent linkage of the enzyme to the natural protein self-assembly system or (ii) by construction of a fusion protein comprising the S-layer protein and the laccase. Combined quartz crystal microbalance with dissipation monitoring (QCM-D) and electrochemical measurements revealed that rLac immobilized on the SbpA lattice had an approximately two-fold higher enzymatic activity compared to that obtained with the fusion protein.

9:35 Integrating High-Throughput Biotherapeutic and Protein Reagent Expression

David Lee, Ph.D., Principal Scientist, UCB Pharma

10:05 Sponsored Presentations (*Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com*)

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

Case Studies In Success

11:05 Diversity is Key: Approaches to Tackle the Expression and Purification of Complex Human Proteins to Drive Early Drug Discovery

Rene Assenberg, Ph.D., Investigator III, Expertise Platform Proteases (EPP), Novartis Pharma AG/NIBB

Production of complex, human proteins to support early drug discovery efforts remains a significant challenge. In our environment we routinely deal with proteins of a highly diverse nature ranging from cytosolic to membrane-bound to secreted protein, either in isolation or in complex with other proteins. To achieve the greatest chances of success, we utilize an equally diverse range of protein expression and purification tools, which will be the focus of this talk.

11:35 Recombinant Production in *E. coli* and Characterization of the Neurotrophic Growth Factor Artemin

Stefan Masure, Ph.D., Principal Research Scientist, Protein Production Group, Janssen Research & Development

A case study will be presented on the recombinant production and detailed characterization of Artemin, a neurotrophic growth factor from the GDNF-family considered as a possible therapeutic agent for neuropathic pain. Efficient *E. coli* expression and purification protocols were developed and the purified protein was characterized in terms of purity, cellular activity, dimerization and stability. The produced protein was suitable for *in vitro* and *in vivo* testing of Artemin's biological activity.

12:05 Reconstituted Nonribosomal Production of the Peptide Antibiotic Valinomycin in the Heterologous Host *Escherichia coli*

Jennifer Jaitzig, Ph.D., Researcher, Department of Biotechnology, Technische Universität Berlin

Nonribosomal peptide synthetases (NRPSs) are multi-functional mega-enzymes that produce a wide range of pharmaceutically relevant peptides in bacteria and fungi. The heterologous expression of NRPSs in a robust host like *Escherichia coli* is a promising approach to make relevant NRPs more accessible. We conducted a rational expression screening in multi-well plates to produce the 655 kDa heterodimeric valinomycin synthetase (VImSyn) from *Streptomyces tsusimaensis* in a soluble form in *E. coli*. VImSyn was purified and its activity was confirmed *in vitro*. Finally, the biosynthesis of valinomycin, a cyclodepsipeptide with reported antifungal, antibacterial and antiviral activities could be fully reconstituted in an engineered *E. coli* strain with the *Bacillus subtilis* phosphopantetheinyltransferase gene *sfp* genomically integrated.

12:35 Close of Part One: Enhancing Expression and Achieving Higher Throughput

"Excellent overview of the most important topics and challenges in the field of next generation protein therapeutics"

Dragan Grabulovski, CSO, Covagen

"A well thought out program – great length and number of talks."

Katherine Vousden, Senior Scientist, Antibody Discovery and Protein Engineering, MedImmune

Part Two: Solving Difficult Protein Problems

7–8 November 2012

WEDNESDAY 7 NOVEMBER**12:30 – 14:00 Conference Registration for Part Two****14:00 Chairperson's Remarks****Membrane Proteins****» KEYNOTE PRESENTATION****14:05 Nanobodies as Tools for the Structural and Functional Investigation of GPCR Transmembrane Signaling**

Jan Steyaert, Ph.D., Executive Director, Department of Molecular and Cellular Interactions, Vlaams Instituut voor Biotechnologie & Structural Biology Brussels, Vrije Universiteit Brussel

We generated Nanobodies that stabilize transient functional conformations of the human β_2 adrenergic receptor. Some Nanobodies that faithfully mimic G protein binding were used to crystallize active agonist-bound states of this GPCR. Other nanobodies that stabilize the $\beta_2\text{AR}\cdot\text{Gs}$ complex were instrumental to purify and obtain the crystal structure of this complex, providing the first view of transmembrane signaling by a GPCR.

14:35 Optimization of Expression and Purification of the Feline and Primate Foamy Virus Transmembrane Envelope Proteins Using a 96 Deep Well Screen

Michael Mühle, Ph.D., Researcher, Robert Koch Institute

High level overexpression of a recombinant target protein in *E. coli* is a prerequisite to minimize efforts in subsequent large scale purification steps. We developed a 96 well based screening method for rapid detection of suitable expression conditions and applied it for the production of two viral transmembrane envelope proteins to quickly determine conditions for an about 20-fold increased production.

15:05 CaptureSelect: Unique Affinity Products for Purification and Detection of All Antibody Formats

Frank Detmers, Ph.D., Director, Ligand Application, BAC BV

This presentation will focus on CaptureSelect IgG-CH1, a novel affinity resin, using a ligand directed towards the CH1 domain of antibodies and antibody fragments, offering a true platform for purification of all human IgG's and Fab fragments thereof, irrespective of subclass and without co-purification of free light chains. Analytical assays based on HPLC column formats and label free detection systems for QC will also be presented.

15:20 Sponsored Presentation (*Opportunity available, please contact Carol Dinerstein, Dinerstein@healthtech.com*)

15:35 Refreshment Break in the Exhibit Hall with Poster Viewing**16:15 Predictable Protein Expression in Mammalian Cells by Exploitation of Defined Chromosomal Loci**

Dagmar Wirth, Ph.D., Department of Gene Regulation and Differentiation, HZI- Helmholtz Centre for Infection Research

The expression levels of transgenes in mammalian cells are affected by the impact of neighbouring chromosomal elements. To achieve predictable protein expression we exploited methods for targeting expression cassettes to specific chromosomal sites via recombinase mediated cassette exchange or employing bacterial artificial chromosomes, respectively. We show that defined combinations of integration sites and vector designs are required for rational exploitation of given chromosomal sites.

16:45 Improving Protein Stability and Function through Disulfide Engineering: A Computational Approach

Alan Dombkowski, Ph.D., Assistant Professor, Division of Clinical Pharmacology and Toxicology, Department of Pediatrics, Wayne State University School of Medicine

Protein stability and function can be enhanced in a wide range of biomedical and industrial applications through disulfide bond engineering. Computational methods have proven to be effective for designing strategically placed disulfides. We will discuss structural considerations for successful use of this technology, recent advances in our computational method for rational disulfide design, and applications.

17:15 Problem Solving Roundtable Discussions**Table 7: Nanobodies as Tools for Structural Studies of Membrane Proteins**

Moderator: Jan Steyaert, Ph.D., Executive Director, Department of Molecular and Cellular Interactions, Vlaams Instituut voor Biotechnologie & Structural Biology Brussels, Vrije Universiteit Brussel

Table 8: Predicting Protein Expression in Mammalian Cells Using Defined Chromosomal Loci

Moderator: Dagmar Wirth, Ph.D., Department of Gene Regulation and Differentiation, HZI- Helmholtz Centre for Infection Research

Table 9: Cell Free Expression of Mammalian Proteins: Strategies for Success

Moderator: Frank Bernhard, Ph.D., Centre for Biomolecular Magnetic Resonance, Institute for Biophysical Chemistry, University of Frankfurt/Main

Table 10: Using Bacterial Artificial Chromosomes to Enhance Protein Expression in Mammalian Cell Lines

Moderator: Emilio Casanova, Ph.D., Researcher, Ludwig Boltzmann Institute for Cancer Research, Vienna

Table 11: Choosing the Right Host for Expression of Membrane Proteins

Moderator: Annie Barrand-Frelet, Ph.D., CNRS, CEA-France

18:15 – 19:15 Reception in the Exhibit Hall with Poster Viewing**THURSDAY, 8 NOVEMBER**

7:30 Breakfast Presentation (*Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com*) **or Morning Coffee**

8:30 Chairperson's Remarks**Mammalian and Cell-Free Strategies****8:35 Strategies for the Cell-Free Expression of Membrane Proteins**

Frank Bernhard, Ph.D., Centre for Biomolecular Magnetic Resonance, Institute for Biophysical Chemistry, University of Frankfurt/Main

9:05 Membrane Protein Synthesis with Batch-Mode Cell-Free Protein Synthesis: Facilitated Screening and Efficient Sample Production for NMR

Anders Pedersen, Ph.D., Swedish NMR Centre, University of Gothenburg

Batch-mode cell-free protein synthesis facilitates sample condition screening of membrane proteins, in particular detergent choice for

co-translational solubilization. More than 90% of tested proteins can be expressed in solubilized form. After finding the optimal expression condition for a given membrane protein, isotopic labeling for NMR spectroscopy applications is straightforward and can be accomplished with unparalleled efficiency and specificity.

9:35 The Use of Bacterial Artificial Chromosomes for Recombinant Protein Production in Mammalian Cell Lines

Emilio Casanova, Ph.D., Researcher, Ludwig Boltzmann Institute for Cancer Research, Vienna

Bacterial Artificial Chromosomes (BACs) are vectors derived from the *E. coli* F factor with a cloning capacity of up to 400Kb. Due to their large cloning capacity, BACs can accommodate a whole mammalian locus including all the transcriptional regulatory elements. Expression from BACs-based vectors is not affected by the genomic integration site, BACs-based vectors confer copy-number dependent expression and expression is stable over time. Therefore, BACs should be useful as expression vectors in the field of recombinant protein production in mammalian cells lines.

10:05 Recent Advances in the Integrated Measurements of Protein Size, Structure and Formulation Viscosity

E. Neil Lewis, Ph.D., CTO, Malvern Instruments

The non-invasive and non-destructive determination of numerous physicochemical properties of protein therapeutics is critical for developing optimal formulation conditions to enhance product efficacy, stability and manufacturability. To meet this demand, analytical tools that rapidly measure formulation viscosity, protein structure and hydrodynamic size over formulation concentration ranges on sample volumes < 10 µl are required. In this talk, we describe recent innovations that enable integrated, automated measurements of protein therapeutic formulations on minimal sample volumes.

10:35 Coffee Break in the Exhibit Hall with Poster Viewing

Solubility, Aggregation and Immunogenicity

11:05 On the Role of Aggregates and Particles in Protein Immunogenicity

Wim Jiskoot, Ph.D., Professor, Drug Delivery Technology, University of Leiden

In this presentation I will discuss product-related risk factors for protein immunogenicity (in particular aggregates, submicron particles and subvisible particles), show clinical and preclinical data illustrating the impact of the formulation on protein immunogenicity, and the use of animal models to assess which of these factors do or do not affect immunogenicity.

11:35 Strategies for Bacterial Expression of Protein-Peptide Complexes: Application to Solubilization of Papillomavirus E6

Sebastian Charbonnier, Ph.D., Researcher, Equipe Oncoprotéines

The papillomavirus oncoprotein E6 induces cervical cancers and had resisted structural analysis for more than 20 years. By combining different strategies including production optimisation, mutagenesis, domain phasing, fusion to the highly soluble Maltose-Binding-Protein (MBP) and/or to E6-binding peptides, we ultimately managed to solve several high-resolution structures of E6 proteins in isolation or in complex with target proteins.

12:05 Increasing Yield, Biological Activity and Reducing Cost of Recombinant Protein Production Using Novel Conformases

M. Raafat El-Gewely, Ph.D. Professor, Institute of Medical Biology, University of Tromsø

I will discuss the fact that primary structure is not alone in affecting the proper tertiary structure and the functionality of proteins. Besides improving the environmental conditions for protein expression, we have improved the molecular and genetic

environment in the host (initially *E. coli*) by our novel conformases / foldases (patents pending) which can help reduce the impact of the bottle-neck problems in the recombinant field and drug discovery. This technology has helped produce proteins that were not possible before and is suitably poised to be used in the biosimilar and biobetter field.

12:35 Luncheon Presentations or Lunch on Your Own

(Opportunities available, please contact Carol Dinerstein, Dinerstein@healthtech.com)

The Future of Expression: Advanced Technologies

14:00 Chairperson's Remarks

14:05 Heterologous Expression of Membrane Proteins: Choosing the Appropriate Host

Annie Barrand-Frelet, Ph.D., CNRS, CEA-France

The major bottleneck for characterization of a membrane protein is its overexpression. Expression of 20 membrane proteins of various origins, functions and topologies, has been tested in different prokaryotic and eukaryotic systems. First, genes were cloned using the Gateway technology. Finally, 17 of the 20 proteins were produced at adequate yields for functional and/or structural studies.

14:35 Inducible Expression of G Protein-Coupled Receptors in Transfected Cells

M. Emmanuel Hermans, Ph.D., Institute of Neurosciences, Université Catholique de Louvain

Commercially available systems offer the possibility to induce graded levels of receptor expression in the experimental model, or to induce an abrupt downregulation of receptor expression during the maintenance of the cell-line. This presentation provides an overview of the different systems available and provides methods for the generation and validation of stably transfected cell-lines expressing the GPCR of choice.

15:05 Native Source Protein Purification for Crystallography

Chloe Zubieta, Ph.D., European Molecular Biology Laboratory, Grenoble, France

With improvements in chromatographic methods, the advent of microfluidic technology, and the availability of high flux beamlines with smaller beam sizes, microgram quantities of purified protein can be used for successful crystallographic characterization. Strategies for protein fractionation and purification from native sources will be discussed with a focus on downstream crystallographic applications. Case studies in model and non-model systems will be presented.

15:35 Refreshment Break in the Exhibit Hall with Poster Viewing

16:00 Transient Gene Expression in HEK293 and Vero Cells Immobilised on Microcarriers

Lukas Fliedl, Ph.D., Junior Scientist, Institute of Applied Microbiology, Vienna, Austria

In this work HEK/EBNA cells were grown and transfected on microcarriers. Cell immobilisation allows easy media exchange after sedimentation. The transfection method was optimized regarding polyethylenimine (PEI) concentration, optimal DNA:PEI ratio, type of PEI, incubation time and polyplex formation time. Transfection efficiencies of up to 33% with pCEP4 and 98% with pMAX were reached. Additionally immobilisation on microcarriers was used to retain the cells during cultivation, thus allowing media replacement and prolonging cultivation time from one to two weeks with continuous expression of the recombinant protein.

16:30 Panel Discussion: Where Will We Be in 5 Years in Protein Expression?

17:00 Close of Conference

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Monday, 5 November

SC1: From Understanding of Aggregation to Devising of Prevention Strategies	SC3: Engineering of Bi-Specific Antibodies
SC2: Measures to Enhance Half-Life and Stability	

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Advance Registration Deadline until 28 September	€1450	€725	
Registrations after 28 September and on-site	€1625	€795	€345

Schedule at a Glance

	Novel Antibody Constructs and Alternative Scaffolds	Protein Expression
6-7 November	Creative Engineering, Delivery Technologies and Targeting	Enhancing Expression and Achieving Higher Throughput
7-8 November	Enhanced Product Properties and Therapeutic Application	Solving Difficult Protein Problems

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